

RIBES

PHPS 14

NS

Special requirements for Ribes (Black currants, red currants, white currants and gooseberries) plants to qualify as nuclear stock

Eligible material	Any new or established variety or candidate material of potential new varieties can be entered. The progeny of nuclear stock is eligible as parent material to produce Foundation grade cuttings or for planting stoolbeds for entry at Super Elite.
Growing conditions	<p>Nuclear stock plants must have been maintained in a suitably designed insect-proof gauzehouse containing only Ribes nuclear stock plants.</p> <p>Strict precautions should be taken to prevent the introduction of any pest or disease especially those listed in Annex 1. Quality pests and diseases such as two spotted (red spider) mite or powdery mildew should be kept at as low a level as practically possible.</p> <p>All mother plants must be grown singly in sterilised growing medium and in individually labelled containers.</p>
Pests and diseases	<p>At least every two years the mother plants must have been individually tested and found free from the diseases listed in Annex 1 using the indicator plants or test methods described.</p> <p>New plants to be entered into the nuclear stock house must have been tested and found free from all the diseases listed in Annex 1.</p> <p>Any plants found to be infected with the diseases listed in Annex 1 or exhibiting suspicious symptoms should be removed immediately.</p>
Documentation	<p>The Director or other person responsible for the production of the plants must provide documentary evidence to show that the material has been produced under the conditions described above and that all the necessary tests were carried out and no evidence of infection was found.</p> <p>This evidence must be provided to the purchaser of the nuclear stock material before it can be used as parent material to produce Foundation cuttings or planted to give a Super Elite stoolbed.</p>
Reference for further details	EPPO 1994 Certification scheme for Ribes , EPPO Bulletin 24, 857–864. 1994. Martin RR (Ed) 2004. Recommended procedures for detection of viruses in small fruit crops. Proceedings of Tenth International Symposium on Small Fruit Virus Diseases. Acta Hort. 656:199-207, 2004.



Disease	Test method and indicator plant
<p>Black currant (<i>R. nigrum</i>):</p> <p>Reversion agent</p> <p>Vein banding agent</p> <p>Strawberry latent ringspot nepovirus Arabis mosaic nepovirus Cucumber mosaic cucumovirus</p>	<p>Graft inoculation to black currant cv. Baldwin or Ojebyn</p> <p>Graft inoculation to red currant cv. Jonkeer van Tets, black currant cv. Amos Black, hybrid Ribes EM 1385/81 or 1385/90 or gooseberry cv. Leveller</p> <p>— Mechanical inoculation to test plants of <i>Chenopodium quinoa</i>, <i>Cucumis sativus</i> or <i>Nicotiana clevelandii</i> as applicable. The nepoviruses should be confirmed serologically.</p>
<p>Red currant (<i>R. rubrum</i>) and White currant (<i>R. sylvestre</i>):</p> <p>Reversion agent</p> <p>Vein banding agent</p> <p>Strawberry latent ringspot nepovirus Raspberry ringspot nepovirus Arabis mosaic nepovirus</p>	<p>Graft inoculation to black currant cv. Baldwin or Ojebyn</p> <p>Graft inoculation to red currant cv. Jonker van Tets, black currant cv. Amos Black, hybrid Ribes EM 1385/81 or 1385/90 or gooseberry cv. Leveller</p> <p>— Mechanical inoculation to test plants of <i>Chenopodium quinoa</i>, <i>Cucumis sativus</i> or <i>Nicotiana clevelandii</i> as applicable. They should be confirmed serologically.</p>
<p>Gooseberry (<i>R. uva-crispa</i>):</p> <p>Vein banding agent</p> <p>Arabis mosaic nepovirus Raspberry ringspot nepovirus</p>	<p>Graft inoculation to red currant cv. Jonkeer van Tets, black currant cv. Amos Black, hybrid Ribes EM 1385/81 or 1385/90 or gooseberry cv. Leveller</p> <p>— Mechanical inoculation to test plants of <i>Chenopodium quinoa</i>. They should be confirmed serologically.</p>

Notes:

- For graft inoculation tests, one indicator plant should be used for each virus being tested for. Tests for reversion should be observed for two years and tests on several shoots from large bushes are necessary because of the uneven distribution of this agent in the plant.
For other pathogens tests should be continued until the following spring.
Alternative indicator plants may be acceptable depending on the country of origin of the material, Fera should be consulted in such cases.
- For mechanical inoculation tests on herbaceous indicators, plants should be observed for up to 4 weeks. Identification of specific viruses will require serological tests applied to extracts from the herbaceous indicators.